

INTERCONVERSION OF GIBBERELLIN A₁ TO GIBBERELLIN A₈ IN SEEDLINGS OF DWARF *ORYZA SATIVA*

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Abstract—[³H]Gibberellin A₁ ([³H]GA₁) applied to seedlings of dwarf rice (*Oryza sativa* L. cv. Tanginbozu) was metabolized to GA₈. Identification of GA₈ was made by gas-liquid radiochromatography using three liquid stationary phases.

RECENTLY, Nadeau and Rappaport¹ reported the interconversion of [³H]GA₁ into GA₈ and GA₈-glucoside in seeds of *Phaseolus vulgaris*. Railton and Wareing² also studied the metabolism of [³H]GA₁ in potato leaves and although the single acidic, radioactive conversion product they detected was not identified, its chromatographic and electrophoretic properties were consistent with those of GA₈. GA₄ has previously been shown to be converted to GA₁ in shoots of dwarf rice.³ We now report the conversion of [³H]GA₁ into GA₈ in shoots of the same species.

Tritium-labelled GA₁ (sp.act. 420 mCi/mmol) was applied (74 μCi total) in 95% ethanol to shoots of 25 dwarf rice plants. Twenty-four hr after application, shoots were excised from the seeds and roots, surface-washed in absolute methanol and ground in a mortar at 0° with 80% methanol and acid washed sand. The methanol was removed *in vacuo* at 35° and the residual aqueous phase adjusted to pH 9.0 by the addition of an equal volume of 0.5 M phosphate buffer. The buffer phase was washed 4 times at pH 9.0 with equal volumes of diethyl ether (total radioactivity 0.08×10^6 cpm) and then adjusted to pH 3.0 and extracted 4 times with ethyl acetate (17.0×10^6 cpm) and 3 times with *n*-butanol (0.5×10^6 cpm). The residual buffer contained 0.04×10^6 cpm. All counts were adjusted for quenching but not for counting efficiency.

The acidic, ethyl acetate fraction was chromatographed on a silica gel partition column^{4,5} and eluted with increasing amounts of ethyl acetate in *n*-hexane. Fractions were combined according to counts obtained from direct scintillation spectrometry using Bray's scintillant.⁶ The trimethylsilyl ethers of the methyl esters (TMSMe derivatives) of the combined fractions were analyzed by gas-liquid radiochromatography (GLRC) on 3 column liquid phases, 2% SE30, 2% QF1 and 1% XE60. Conditions for GLRC were identical to those described previously.^{3,7} Combined fractions 19–22 contained one significant peak with retention times

¹ NADEAU, R. and RAPPAPORT, L. (1972) *Phytochemistry* **11**, 1611.

² RAILTON, I. D. and WAREING, P. F. (1973) *Physiol. Plant.* **28**, 127.

³ DURLEY, R. C. and PHARIS, R. P. (1973) *Planta (Berl.)* **109**, 357.

⁴ POWELL, L. E. and TAUTVYDAS, K. J. (1967) *Nature (London)* **213**, 292.

⁵ DURLEY, R. C., CROZIER, A., PHARIS, R. P. and MCLAUGHLIN, G. E. (1972) *Phytochemistry* **11**, 3029.

⁶ BRAY, G. A. (1960) *Anal. Biochem.* **1**, 279.

⁷ DURLEY, R. C., RAILTON, I. D. and PHARIS, R. P. (1973) *Phytochemistry* **12**, 1609.

identical to those of TMSMe GA₈ standard (2% SE30, 25.4 min; 2% QF1, 17.4 min; 1% XE60, 17.5 min). Combined fractions 14–18 also contained a single radioactive peak, with identical retention times to those of TMSMe GA₁ standard (2% SE30, 15.4 min; 2% QF1, 14.0 min; 1% XE60, 15.3 min). Combined fractions 23 and 24 contained an unknown radioactive compound exhibiting long retention times on GLRC (2% SE30, 33.0 min; 2% QF1, 56.0 min; 1% XE60, 54.0 min).

The total radioactivity associated with each of the compounds was as follows: GA₈, 1.9×10^6 dpm; GA₁, 4.7×10^7 dpm; unknown compound, 2.4×10^6 dpm. The incorporation of GA₁ to GA₈ was 1.2% of the [³H]GA₁ applied to the tissue.

The results reported here further confirm that GA₁ is preferentially hydroxylated at C-2 to produce GA₈ in higher plants and that such a conversion does occur in vegetative tissue.

Attempts were made to detect the presence of GA₈-glucoside in extracts of rice shoots. The *n*-butanol fraction was adjusted to neutral pH with saturated sodium bicarbonate solution, stored at -20° and subsequently reduced to dryness *in vacuo*. The residue was taken up in a minimum volume of distilled water and desalted on a column of charcoal (1.5 g)–celite (3 g) eluted with water. Radioactivity was removed from the column by washing with absolute methanol and derivatives of the residue from this fraction for GLRC were prepared as described previously.^{3,7} Trace amounts of the free acid of GA₈ were present but the glucoside could not be detected. Recently, Nadeau and Rappaport (personal communication) have detected by co-chromatography using TLC the presence of [³H]GA₈-glucoside after feeding [³H]GA₁ to vegetative tissue of maize.

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